Review

# Some Aspects of Speciation of Mercury in a Water Environment

L. Boszke\*, G. Głosińska\*, J. Siepak \*\*

 \* Department of Environment of Protection, Collegium Polonicum, A. Mickiewicz University, Kosciuszki 1, 69-100 Słubice, Poland, e-mail: boszke@euv-frankfurt-o.de.
 \* Department of Water and Soil Analysis, Faculty of Chemistry, A. Mickiewicz University, Drzymały 24, 60-613 Poznań, Poland, e-mail: zawig@amu.edu.pl.

> Received: 16 January, 2002 Accepted: 1 March, 2002

## Abstract

Extreme toxicity of some species of mercury, the ability of this element to bioaccumulate in particular in fish meat, and the known cases of lethal poisoning by mercury have drawn particular attention to this element's presence in the natural environment. Due to the relatively long time of its presence in the air, elemental mercury can be transported over large distances, hence the presence of mercury of anthropogenic origin is detected practically all over the world. Apart from the elemental mercury. The main species of mercury in water are Hg(II) and mercury-organic species, in particular methylmercury. The latter undergoes strong bioaccumulation in living organisms and concentration in the trophic chains. That is why the relative concentration of mercury in organisms, access to oxygen, illumination, influenced by biotic and abiotic factors such as the activity of microogranisms, access to oxygen, illumination, temperature, pH and others. Despite intense studies, full and reliable recognition of the ecological and health effects of pollution by this toxic metal is still impossible. The aim of this paper is to present the problems related to speciation of mercury, and describe some conversion and migration processes of mercury in the water environment.

Keywords: mercury, concentration, speciation, water, environmental transformation, fluxes.

## Introduction

Mercury was one of the first elements whose speciation has been studied. The main incentive of the studies was the serious poisoning by mercury which polluted the waters of Minamata Bay, Japan, where it occurred as methylmercury and dimethylmercury [1, 2]. The phenomena pointed not only to different toxicities of different compounds of the same element but also to the possibility of conversions of these compounds in the environment, in this case to mercury methylation by microogranisms. The mercury species occurring in the environment can be divided into volatile  $(Hg^{\circ}, (CH_3)_2Hg)$ , easily water soluble  $(Hg^{2+}, HgCl_2, Hg(NO_3)_2)$  and hardly soluble organic complexes  $(CH_3Hg^+, CH_3HgS)$ , [3, 4]. Mercury can form compounds not only with a methyl group but also with other alkyl groups [5, 6]. Distribution of mercury compounds depends to a significant degree on red-ox conditions. In the oxidising conditions the dominant forms are  $Hg^{2+}, Hg_2^{2+}$ , etc., while in the reducing conditions the dominant are sulphur-mercury compounds  $HgS, HgS_2^{2-}$ ,  $CH_3HgS^-$ , etc., and in the intermediate conditions the most often met are alkyl mercury compounds [3, 7]. Mercury species occurring in the natural environment can be also divided according to their reactivity. The most reac-

Correspondence to: Prof. Jerzy Siepak

Table 1. Concentrations of mercury (ng/L) in water from various parts of the world.

	Hg (total)	Hg (total dissolved)	DGM	Hg (reactive)	MeHg	Hg (particulate)	Refe- rences
		Sea Wat	ter				
Baltic Sea	0.6 ± 0.2				~20% Hg(total)		[12]
Gulf of Gdańsk	277-630						[11]
Gulf of Finland	10-140	alaan	230	apto	mark ins	an2	[11]
Bothnian Bay	2-40	moory	0.10	0333	d care as	48.010	[11]
Gulf of Riga	10-40	Ernst	1918	N R	TT I		[11]
Matsalu Bay (Baltic Sea)	5-130	1	1.4.4.4.4.4				[11]
North Sea	0.56 ± 0.24						[24]
North Sea	0.5-200					7	[73]
Mediterranen Sea (Tunisia)	990-27060	* rolzmie	10.0.1	30526			[74]
Northeastern Atlantic	2.7						[23]
Sepetiba Bay (Brazil)		0.12-0.67	0.032-0.082	0.11-0.36	< 0.020-0.55	alight the	[25]
		River Wa	ter		L DS DUCI LA		
Ebro Delta (Spain)	1-18	outra lin	Guo rout/	6.10000	spall States		[75]
Gösku Delta (Turkey)	156-1502			11.00			[76]
Rivers inflowing to Onondaga Lake (USA)	1.0-104						[41]
Rivers inflowing to Narragansett Lake (USA)	7.2-69.2						[77]
Rivers in Wisconsin-industrial area (USA)	1-43						[78]
Tapajos River (Brazil)	0.28-13.3		1				[79]
St. Lawrence River (Canada)	0.08-2.0						[80]
Indrija River (Slovenia)	2.77-322	0.43-39.23			< 4.52		[13]
Madeira River (Brazil)	2.25-20.05	0.28-8.06			0.61-1.825		[13]
Carson River (USA)	4-7585	0.2-88			0.3-7.2		[13]
Rivers inflowing to Michigan Lake	0.98-21.9	-			0.050-0.360		[16]
St. Lawrence Estuary	0.4-1.5			< 20% Hg(total)			[81]
Various estuaries in Florida (USA)	3-7.4				<0.002-2.3		[82]
Chattahoochee River (USA)		0.12-3.174				8-18*	[42]
Various streams in Southern USA	< 2100				< 0.8		[83]
Gironde Estuary (France)	< 250	< 3.0			< 1.8	< 60*	[84]
Scheld Estuary (Belgium/Netherlands)	< 150	< 2.4			< 1.4	< 24*	[84]
Rhine Estuary (Netherlands)	< 20	< 2.0			< 0.1	< 10*	[84]
		Lake Wa	ter				
Lake Michigan (USA)	0.56-182						[85]
Lake Michigan (USA)	$0.32\pm0.05$						[86]
Lake Michigan (USA)	0.32				0.005-0.042**	0.64-0.16	[87]
Lake Michigan (USA)	0.30 (0.2-0.4)	0.204 (0.11-0.31)			0.0075 (0.0075-0.014)	60	[88]
Lake Michigan (USA)					0.0047 (0.0054-0.011)**		[88]
Various water reservoirs (Canada)						200-600*	[89]
	- T	Interstitial	Water				
Sepetiba Bay (Brazil)			< 2.6	0.2-32.2	0-246		[90]
St. Lawrence Estuary	3.3-15.6			< 30% Hg (total)	1.0-2.8		[81]

\* ng/g; \*\*measurements "dissolved" fraction of Hg; Hg(total) - total mercury; DGM-dissolved gaseous mercury; Hg(reactive) - "reactive" mercury; MeHg - monomethylmercury; Hg(particulate) - mercury bound with particles. tive species are:  $Hg^{2+}$ ,  $HgX^+$ ,  $HgX_2$ ,  $HgX_3^ HgX_3^{2-}$ (where X = OH<sup>-</sup>, CI<sup>-</sup>, or Br), Hg° bound to aerosol molecules and Hg<sup>2+</sup> bound to organic acids (water-soluble). The unreactive species include CH<sub>3</sub>Hg<sup>+</sup>, CH<sub>3</sub>HgCl, CH<sub>3</sub>HgOH and other mercury-organic compounds, Hg(CN)<sub>2</sub>, HgS and Hg<sup>2+</sup> bound to sulphur in the humic substances [8]. Certain species belonging to the chemically unreactive ones (e.g. CH<sub>3</sub>Hg<sup>+</sup>) are bioavailable and can undergo bioaccumulation and biomagnification. Moreover, some chemically unreactive species can be converted to the reactive ones through biochemical transformations [4, 9, 10].

## Speciation of Mercury in Hydro-Ecosystems

#### Concentrations of Mercury Species in Hydrosystems

In natural waters, especially in marine water, mercury occurs at very low concentrations, which causes many serious problems with their accurate determination. The literature data on the concentration of mercury in natural waters are unreliably diverse not only as a consequence of the natural variation of the species and concentrations of this metal in water (Table 1 and 2), but also because of analytical difficulties. For instance, the report of the Baltic Marine Environmental Protection Commission in Helsinki [11] says that the concentration of mercury in Gdansk Bay reaches from 227 to 630 ng/L, which is a value much higher than that reported for any other Baltic region. However, according to recent data the concentration of mercury in different regions of the Baltic Sea, Gdansk Bay included, is much lower  $-0.6 \pm 0.2$  ng/L [12]. It has been estimated that the natural concentration of mercury in marine waters varies from 0.5 to 3.0 ng/L, while in estuaries and precipitation it varies from 2.0 to 15 ng/L [9]. In strongly polluted waters the concentration of total mercury reaches a few tens  $\mu$ g/L [4, 13, 14].

Reliable determination of methylmercury concentration in water is even more difficult than that of total mercury. It is estimated that the contribution of methylmercury in the concentration of total mercury in natural non-polluted waters is 3-6% in sea water and 26-53% in fresh water [15]. The data on the concentration of methylmercury in natural waters (and its contribution in the total mercury concentration) are characterised by significant ambiguity (Table 1 and 2). For instance, the contribution of methylmercury in the concentration of total mercury in unfiltered water from the non-polluted river Pere Merquette was determined as 5% [16], and was of the same order of magnitude as that determined for the water from Onondaga lake [17]. The contribution of methylmercury in the concentration of total mercury in the pore water from the same lake was 37% [17] and that in the pore water from the polluted Saquenay Fjord in Canada was smaller than 1% [18].

In surface waters mercury does not occur in the form of free  $Hg^{2+}$  ions but as a mixture of compounds in which mercury form a hydroxy- and chloro-complexes in proportions depending on pH and the concentration of chloride ions (Fig. 1 and 2). In marine water the dominant mercury compounds are chlorine complexes. The occurrence and migration of different mercury species in

Precipitation Water	Hg(total)	Hg(reactive)	MeHg	References
Gardsjon Lake – (Sweden)	*		0.1-0.66	[91]
Halle/Leipzig/Bitterfold (Germany)	460	30		[92]
Mace Head (Ireland)	5.1-37.8	3.7-17.3		[92]
Langenbrugge - 200 km from industrial area (Germany)	45.7-58	3.1-5.7		[92]
Sylt Island – North Sea (Germany)	6.1-18.3	0.6-1.3		[92]
Zingst Penisula - Batic (Germany)	20-111	8.2-31.2		[92]
Scandinavia	5-40			[93]
Experimental Lakes Area - Ontario (Canada)	0.95-9.3		0.010-0.18	[94]
Quebec (Canada)	7.0			[95]
Florida (USA)	5-113			[96]
Chesaepake Bay (USA)	2-8		1	[44]
Michigan (USA)	1.2-60			[97]
Florida (USA)	9-26		< 0.005 - 0.020	[98]
Lake Superior (USA)	9			[99]
Wisconsin (USA)	8			[100]
Wisconsin (USA)	2.7-150			[101]
Wisconsin (USA)	$10.5 \pm 4.8$		$0.16 \pm 0.07$	[102]
Maine (USA)	8.9 ± 3.8		$0.17 \pm 0.08$	[103]
Hoyama (Japan)	5-187			[104]

Table 2. Concentrations of mercury (ng/L) in precipitation water from various parts of the world.

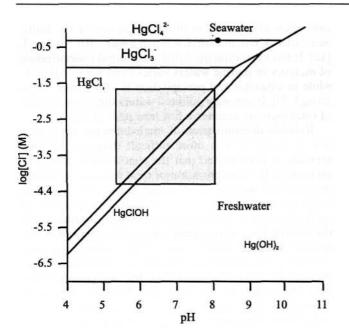


Fig. 1. Dominance diagram of hydroxo- and chloro-complexes of Hg(II) as a function of pH and chloride concentrations [10].

water environment depends also to a great degree on the red-ox conditions and the content of dissolved organic carbon (DOC). In water of low pH, the dominant are the soluble mercury compounds, such as  $HgCl_2$  and  $CH_3Hg^{2+}$ , whereas in mild alkaline conditions  $Hg^{\circ}$  and  $(CH_3)_2Hg$  dominate. In the water of oxidative properties the dominant compounds are  $HgCl_4^{2-}$  and  $HgOH^+$ , while in reductive conditions it is  $CH_3HgS^-$  and  $HgS_2^-$ , and in variable conditions we most often find  $CH_3HgCl$  and  $CH_3Hg^{2+}$  [3, 9, 10, 19-21]. In polluted waters, mercury compounds with phenyl groups were noted [21]. It has been estimated that the lifetime of mercury in water can last from a few minutes to many years, depending on the type of mercury species [22, 23].

#### Speciation of Mercury in the Suspended Phase

Mercury occurs in water in the dissolved phase as well as in the colloidal and suspended phases. The contribution of mercury in these two phases varies both in time (seasonal changes) and in space (e.g. bathymetric depth), and is also affected by living organisms (phytoplankton and bacteria) forming a kind of organic suspension. For example in the suspension from the North Sea, the contribution of methylmercury to total mercury was -6% [24]. In water from Sepetiba Bay the contribution of mercury in the suspended phase made about 70% of the total mercury concentration in water [25]. Although there is little data on the chemical species of mercury in the suspended phase, it is supposed that the mercury species bound to the organic suspension are dominant [26]. Organic matter, in particular in water rich in humic substances, is surrounded by iron and manganese hydroxides in the colloidal phase. The molecules undergo intense

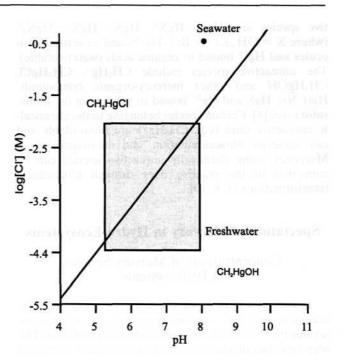


Fig. 2. Dominance diagram of hydroxo- and chloro-complexes of methylmercury as a function of pH and chloride concentrations [10].

aggregation, which is the main mechanism of removal of heavy metals, including mercury, from bulk water to bottom sediments, especially in estuaries [27]. The predominant role of organic matter in migration of mercury in water environment is supported by a strong positive correlation between the concentration of total mercury and the content of organic matter in bottom sediments, established in different parts of the world [28-31]. In Puck Bay, a relatively homogeneous distribution of mercury in bottom sediments, after normalisation of the concentrations and their expression per units of organic matter, unambiguously indicates the sedimenting organic matter as the main source of this element in bottom sediments. At the same time it points to the atmosphere as the main source of this metal reaching the organic matter [31, 32]. The magnitude of the inflow of mercury from different parts of the world to the bottom sediments are given in Table 3. These magnitudes most often correspond to the inflow of mercury through the atmosphere to water reservoirs and illustrate the role of water suspensions in biogeochemical cycle of mercury.

#### Speciation of Mercury in the Dissolved Phase

In the dissolved phase mercury can occur in the form of many species: elemental mercury  $(Hg^{\circ})$  volatile and relatively unreactive, ionic [Hg(II)] in a number of compounds, organic - mainly methylmercury  $(CH_3Hg^+)$  or dimethylmercury  $((CH_3)_2Hg)$ . In strongly stratified water ecosystems elemental mercury is mainly found in surface layers, whereas the total mercury (in dissolved phase and suspended phase) and methylmercury are determined in higher concentrations in the bottom layer [33]. There is

with with a second with a second seco	Inflow of total mercury to bottom sediments	Inflow of MeHg to bottom sediment	References
	Europe		
Batic Sea	44		[105]
Baltic Proper	41	reary =0	[105]
Gulf of Botnian	112		[105]
Gulf of Bothnian	90		[106]
Bothnian Sea (Baltic)	16		[105]
Bothnian Sea (Baltic)	88		[106]
Gulf of Finland	112		[105]
Gulf od Riga	47		[105]
Central Norway	28		vide [107]
	North America		
Onondaga Lake (USA)	8500-13400		[108]
Lake Michigan (USA)	57.4	1.0	[88]
Lake Michigan (USA)	90 ± 52 - 274 ± 238 (23-876)		[45]
Lakes in Wisconsin (USA)	126-305		[109]
Lakes in Midwest USA	34.2		[110]
Fjord Sequenay (Canada)	537		[18]
Imitavik Lake (Canada) – 1980's – 1990's	34 49		[107]

Table 3. Inflow of mercury (ng/m <sup>2</sup>	/day) to bottom sed	iments from different	parts of the word.

also the easily-reducible fraction of mercury (dissolved and suspended phases) undergoing reduction upon treatment by a solution of SnCl<sub>2</sub>. This fraction includes the species easily reducible also in natural conditions and most probably made of inorganic compounds. Investigation of mercury species in vertical water columns revealed an increase in the concentration of divalent gaseous mercury (DGM) in the surface layers relative to that in bottom layer, most probably due to a greater reduction and demethylation in accordance with greater primary production and microbial activity in the surface layers [25]. In the water from Sepetiba Bay the concentration of total reactive mercury in the surface layer varied from 55% to 90% of total soluble mercury, while the organic dissolved Hg made 10% - 83% of total soluble mercury. In general, reactive Hg occurred in higher concentrations in the regions with high concentration of the suspension [25]. In water from the North Sea the contribution of reactive Hg to total Hg was ~30%, while the contribution of mercury in the suspended phase to the concentration of total mercury varied from 13% to 82% [24]. The concentration of total mercury in the dissolved phase in the river Ochleckonee and its estuary (the USA) varied from 0.6 to 6.0 ng/L, while in water from the northern Atlantic - from 0.1 to 0.5 ng/L. The contribution of mercury in the colloidal fraction (>1 kD) in the water from

the estuary was 35% - 87% of total mercury in the dissolved phase (at a low salinity it was 79% - 87%) and in water from the northern Atlantic it was 10% - 50% [34].

#### Speciation of Mercury in Water Catchments

Some amount of divalent mercury is bound to humic acids, estimated to contain 50% - 90% of dissolved organic carbon (DOC). The fraction of divalent mercury bound to DOC has been estimated at 95% [35]. The strong affinity of divalent mercury [Hg(II)] to organic matter plays a very important role in transportation of mercury from catchment to water reservoirs [36-40]. The laboratory tests of Wallschlager et al. [36] proved that soil polluted with mercury and flooded by water from the river Elba released only 1% of mercury contained in the soil upon their treatment with water of pH close to natural. The mercury liberated from the samples was almost completely bound to the humic acids, which points to a large ion-exchange capacity of the humic substances contained in the soil [36]. The magnitude of mercury outflow with water in different parts of the world is given in Table 4. The estimated outflow of mercury with the river water expressed per  $m^2$  of catchment area varies from 2.2 to 66 ng/m<sup>2</sup>/day. Of course, greater loads of the mercury out-

all the second s	Area of catchment (1000 x m <sup>2</sup> )	Outlow of total mercury with river waters	Outflow of MeHg with river waters	References
1	E	irope		
Iso Lake (Finland)	39	4.1-4.7		[111]
15 various catchments (Sweden)		2.2-16		[112]
Gardson Lake (Sweden)	37	6.3-9.6		[91]
Tiveden National Park (Sweden)	14	9.3		[113]
Catchment in South Sweden	50	4.2	0.33	[49]
	North	America		di use nunti
Chesapeake Bay (USA)	16x10 <sup>7</sup>	2.6		[44]
Catchments in Wisconsin (USA)		5-14		[78]
Onondaga Lake (USA)	6.9x10 <sup>5</sup>	14-66		[108]

Table 4. Outflow of mercury with the rivers water  $(ng/m^2/day)$  expressed per m<sup>2</sup> catchment area in various parts of the world.

flow with river waters are observed in industrialised areas - 14-66 ng/m<sup>2</sup>/day than in rural parts - 2.2-6.8 ng/m<sup>2</sup>/day as in the industrialised areas the deposition of atmospheric mercury and the concentration of mercury in waters are usually greater [41, 42]. In general, inflow of total mercury through natural waterways is smaller than the deposition of atmospheric mercury [43]. In Chesapeake Bay [44] and Lake Michigan [45] direct deposition of mercury from the atmosphere is an important source of this element. In these two water reservoirs the contribution of mercury deposited from the atmosphere has been estimated at 50% of all mercury flowing into these reservoirs from all sources. Total mercury of atmospheric origin is strongly bound in the soil and water suspension. It has been estimated that 90% of total mercury coming from the atmosphere is captured in the catchment area [44]. For the take of comparison, the contribution of mercury mass flowing into Onondaga Lake from the catchment area in total mass of mercury flowing into the lake has been estimated as -25% [41]. Unfortunately, information on the behaviour of total mercury cannot be used to infer about the behaviour of methylmercury [16, 39, 40, 43, 46]. In Chesapeake Bay the dominant inflow of methylmercury comes from waterways (25-50 mol/year) and not from the atmosphere (~7 mol/year). The mass of methylmercury deposited from the atmosphere in the Bay catchment area (80 mol/ year) is of a similar order of magnitude as that flowing into the Bay from the waterways. This fact indicates that methylmercury is produced in the catchment area, even if we assume that methylmercury is to a lesser degree bonded to the suspension and soil [44]. When there are no point sources of methylmercury, the main source of this toxic species is precipitation falling directly on the bay water and on the catchment area, the inflow through surface waterways (methylmercury deposited in the catchment area and not captured there, and methylmercury produced in the area), production of methylmercury in the water column or bottom sediments [41,43, 46]. The contribution of these sources is different and depends on the magnitude of atmospheric deposition, type of water reservoir (type of lake) and the size of the wetlands within the catchment area. According to St Louis et al. [43], the wetlands are a source of methylmercury (a net source) and the size of production depends on the type of ground (swamps or peat bogs) and the hydrological conditions there. A smaller production of methylmercury was observed in wetlands of river valleys, where water interaction was restricted to a peat layer and did not reach the mineral substrate. The production of methylmercury was greater at the sites where infiltration water was in contact with the mineral substrate. In the area without wetlands, with aerobic soils, the concentration of methylmercury in the water coming from the waterways is much lower. This finding can be explained by a stronger bonding of methylmercury in these conditions and by greater demethylation taking place on access to oxygen [43]. Therefore, it is obvious that the contribution and distribution of methylmercury depends on the contribution of wetlands in the catchment area. It has been estimated that wet grounds undergoing flooding can produce from 26 to 79 times more methylmercury than the non- flooded grounds [46]. The presence of forests also affects the inflow of mercury to water reservoirs. For instance, in the catchment area of the river Marquette (non-polluted river) forests occupy about 71% of the area and 8% are taken by wetlands. The mean concentration of methylmercury in the water from this river was of the same order of magnitude as that in the river Monistique, whose catchment area is 70% occupied by wetlands [16]. Another factor influencing the presence of methylmercury is the size of precipitation. The greater the amount of the precipitation the greater the outflow of methylmercury from the wetlands [43]. Moreover, the production of methylmercury in natural reservoirs were the same as the amount of methylmercury flown in with the water from the waterways [43]. In artificial water reservoirs the internal production of methylmercury is of greater importance [46]. According to the estimations made by Porvari & Verta [47], the internal production of methylmercury is more important for seepage lakes (up to 67% of total intake) than for drainage lakes (35% of total intake). The same

authors reported that for drainage lakes, whose catchment comprises 10% of wetlands, the contribution of methylmercury flowing in from the catchment is significant and reaches 67% of the total inflow of methylmercury to the lake. For seepage lakes only 5% of methylmercury comes from the catchment area, even when the contribution of wetlands in the catchment area reaches 100% [47].

The effect of the type of catchment on the inflow of methylmercury into the water reservoirs is evidenced by a positive correlation between the concentration of methylmercury in the plankton and the colour of water, the size of the catchment and the contribution of the flooded grounds in the catchment area [48]. The same authors reported a negative correlation between the concentration of methylmercury in the plankton and pH of the water. In brown reservoirs there is in general a higher concentration of methylmercury in fish meat [46]. The study by Petersson et al. [49] also revealed a strong positive correlation between the concentration of methylmercury flowing out of the catchment and the concentration of humic substances.

## Chemistry of Mercury Species in Hydrosystems

## Biotic Methylation of Mercury

Methylation of mercury takes place mainly in bottom sediments and in soils, but it can also take place in water. The process can occur by biotic and abiotic pathways and is affected by many factors such as availability of inorganic Hg(II), activity of microorganisms, red-ox conditions, pH, temperature, salinity, content of organic matter (often described by the colour of water) [4, 9,10]. The biotic methylation takes place mainly in anaerobic conditions, but it can also occur, although more weak, in aerobic conditions [33, 50]. In the biotic methylation of mercury in anaerobic conditions, methylcobolamine acts as a donor of methyl groups [15, 51]. Of key importance in the aerobic process are the sulphate reducing bacteria (SRB) [52-54], with whose involvement in the process can be described as:

$$Hg^{2+} + RCH_3 \twoheadrightarrow CH_3Hg^+ + R'$$
(1)

Monomethylmercury forms easier in the acidic environment, at a relatively high concentration of mercury (Breteler et al., 1981 vide [26]), although Matilainen & Verta [33] reported no effect of pH in the range 4.9 - 6.9 on the methylation of mercury in lake waters. It is believed that in rivers, the optimum temperature of the process is 35°C (Callister & Winfrey, 1986 vide [26]). As far as the effect of salinity is concerned, it has been established that the rate of methylation decreases with increasing salinity, most probably because of the inhibitory influence of chlorine-complexes [55]. The rate of methylation also seems to increase in anaerobic bottom sediments. It has been observed that the amount of methylmercury increased in proportion to the concentration of free sulphide ions (maximum 1.8 mg  $S^{2-}$ / g of sediment). Above this point, the concentration of CH<sub>3</sub>Hg<sup>+</sup> decreased, most probably because of the formation of volatile dimethylmercury (reaction 4) [56]. At a too high concentration of sulphide ions, the concentration of Hg(II) in the solution is too low for methylation to occur and a hardly soluble HgS forms [57]. The mechanisms of mercury methylation in water have not been fully recognised yet, but it is assumed that they involve microorganisms, similarly as in bottom sediments [10]. For instance it is assumed that in lakes methylmercury found in surface layers is transported there by diffusion and currents from the deeper anaerobic layers. In oceans it is supposed that some methylmercury can be formed in aerobic conditions as a result of conversion of dimethylmercury coming from deeper layers. The reactions involving microorganisms lead to the formation not only of monomethylmercury but of dimethylmercury [4, 9]:

$$2CH_3Hg^+ + H_2S \rightarrow (CH_3)_2Hg + HgS + 2H^+$$
 (2)

The rate of formation of these species depends on the concentration of mercury and pH of the environment. Monomethylmercury forms easier in acidic environments, at a relatively high concentration of mercury, while dimethylmercury in neutral or alkaline conditions, at a relatively low concentration of mercury and in the presence of relatively strong complexing reagents such as  $H_2S$  [4, 21]. The rate of monomethylmercury formation is about 6000 times higher than that of dimethylmercury formation, so in the natural environment only 3% of organic mercury occurs as dimethyl species (Regnell, 1990 vide [26]). It is assumed that the formation of dimethylmercury by microorganisms and its liberation to the environment is a detoxication mechanism [51, 52].

#### Abiotic Methylation

The abiotic pathways of methylation can be divided in two groups: those involving and not involving irradiation [15]. In reactions involving irradiation the donors of methyl groups can be acetic acid, propionic acid, methanol and ethanol, whereas the reactions without irradiation include those with methylcobalamine, trasmethylation (methylated tin compounds) and those with humic substances [15]. The role of these agents in methylation of mercury taking place in the natural environment has not been fully determined [57]. Methylkobalamine is a derivative of vitamin B12, in which the terminal ligand CN is substituted with a CH<sub>3</sub> group. A strong correlation has been found between the concentrations of methylmercury and methylcobolamine in bottom sediments [50]. Methylated tin and lead compounds can also be potential reagents in abiotic methylation of mercury, especially in tin and lead-polluted regions [57-59]. The compound CH<sub>3</sub>J, occurring at a relatively high concentration in natural waters and produced by algae, had also been supposed to be involved in direct methylation of mercury. However, experimental data excluded this hypothesis. Nevertheless, CH<sub>3</sub>J can be indirectly involved in methylation of mercury taking part in methylation of tin compounds in water environment [58]. Weber [57] suggested that in marine water CH<sub>3</sub>J can methylate trace amounts of Hg(I) ( $Hg_2^{2+}$ ). From among the methylating agents the most important seems to be humic substances,

$\begin{array}{c} 2-340 \\ 100 \pm 50 \\ 20-360 \\ 180 \pm 60 \\ 100 \pm 30 \\ 400 \pm 240 \\ 3.5-160 \\ 310 \pm 310 \\ 0.74-5.7 \\ 2.8-180 \\ 60-220 \\ 140-190 \\ 30 \pm 10 \\ 30-790 \\ 10-900 \\ < 290 \\ 100-5330 \\ \end{array}$	An vetture inter inter a si the set time a sense of set time a sense of set time a time a set in the set time of the set of set in time a set of set of set in time a set of	[12] [105] [114] [105] [105] [105] [115] [115] [114] [114] [114] [114] [114] [114] [116] [117]
$\begin{array}{c} 100 \pm 50 \\ \hline 20{\text{-}}360 \\ \hline 180 \pm 60 \\ \hline 100 \pm 30 \\ \hline 400 \pm 240 \\ \hline 3.5{\text{-}}160 \\ \hline 310 \pm 310 \\ \hline 0.74{\text{-}}5.7 \\ \hline 2.8{\text{-}}180 \\ \hline 60{\text{-}}220 \\ \hline 140{\text{-}}190 \\ \hline 30 \pm 10 \\ \hline 30{\text{-}}790 \\ \hline 10{\text{-}}900 \\ \hline < 290 \end{array}$		[105] [114] [105] [105] [105] [115] [115] [114] [114] [114] [114] [114] [116]
$\begin{array}{c} 20 - 360 \\ \hline 180 \pm 60 \\ \hline 100 \pm 30 \\ \hline 400 \pm 240 \\ \hline 3.5 - 160 \\ \hline 310 \pm 310 \\ \hline 0.74 - 5.7 \\ \hline 2.8 - 180 \\ \hline 60 - 220 \\ \hline 140 - 190 \\ \hline 30 \pm 10 \\ \hline 30 \pm 10 \\ \hline 30 - 790 \\ \hline 10 - 900 \\ \hline < 290 \end{array}$	Element is a contract of the second of th	[114] [105] [105] [105] [115] [115] [114] [115] [32] [114] [114] [114] [114] [116]
$\begin{array}{c} 180 \pm 60 \\ 100 \pm 30 \\ 400 \pm 240 \\ \hline 3.5-160 \\ \hline 310 \pm 310 \\ \hline 0.74-5.7 \\ \hline 2.8-180 \\ \hline 60-220 \\ \hline 140-190 \\ \hline 30 \pm 10 \\ \hline 30 \pm 10 \\ \hline 30-790 \\ \hline 10-900 \\ \hline < 290 \end{array}$		[105] [105] [105] [115] [114] [115] [32] [114] [114] [114] [114] [116]
$ \begin{array}{r} 100 \pm 30 \\ \hline 400 \pm 240 \\ \hline 3.5-160 \\ \hline 310 \pm 310 \\ 0.74-5.7 \\ \hline 2.8-180 \\ \hline 60-220 \\ \hline 140-190 \\ \hline 30 \pm 10 \\ \hline 30 \pm 10 \\ \hline 30-790 \\ \hline 10-900 \\ \hline < 290 \\ \hline $	<ul> <li>A Jiga Lange Lange</li> <li>A Jiga Lange</li> <li< td=""><td>[105] [105] [115] [114] [114] [115] [32] [114] [114] [114] [114] [116]</td></li<></ul>	[105] [105] [115] [114] [114] [115] [32] [114] [114] [114] [114] [116]
$\begin{array}{r} 400 \pm 240 \\ \hline 3.5-160 \\ \hline 310 \pm 310 \\ \hline 0.74-5.7 \\ \hline 2.8-180 \\ \hline 60-220 \\ \hline 140-190 \\ \hline 30 \pm 10 \\ \hline 30-790 \\ \hline 10-900 \\ \hline < 290 \\ \end{array}$	And a second sec	[105] [115] [114] [115] [32] [114] [114] [114] [114] [116]
$\begin{array}{r} 3.5-160 \\ \hline 310 \pm 310 \\ \hline 0.74-5.7 \\ \hline 2.8-180 \\ \hline 60-220 \\ \hline 140-190 \\ \hline 30 \pm 10 \\ \hline 30-790 \\ \hline 10-900 \\ \hline < 290 \end{array}$		[115] [114] [115] [32] [114] [114] [114] [114] [116]
$\begin{array}{r} 310 \pm 310 \\ 0.74 + 5.7 \\ \hline 2.8 - 180 \\ \hline 60 - 220 \\ \hline 140 - 190 \\ \hline 30 \pm 10 \\ \hline 30 - 790 \\ \hline 10 - 900 \\ \hline < 290 \end{array}$		[114] [115] [32] [114] [114] [114] [114] [116]
$\begin{array}{c} 0.74-5.7\\ \hline 2.8-180\\ \hline 60-220\\ \hline 140-190\\ \hline 30 \pm 10\\ \hline 30-790\\ \hline 10-900\\ \hline < 290\\ \end{array}$		[115] [32] [114] [114] [114] [114] [116]
2.8-180 60-220 140-190 30 ± 10 30-790 10-900 < 290	ortaj (o massi o litti kom n oj (o massi o litti kom n oj (o massi o massi n oj (o massi n oj (o massi o massi o massi o massi o massi n oj (o massi o massi o massi o massi o massi n oj (o massi o massi o massi o massi o massi o massi o massi n oj (o massi o massi n oj (o massi o mas	[32] [114] [114] [114] [114] [116]
60-220           140-190           30 ± 10           30-790           10-900           < 290		[114] [114] [114] [116]
140-190 30 ± 10 30-790 10-900 < 290		[114] [114] [116]
30 ± 10 30-790 10-900 < 290		[114] [116]
30-790 10-900 < 290		[116]
10-900 < 290		[116]
< 290		
100-5330		[118]
		[119]
20-127	0.01-0.053	[66]
38-790		[120]
10-410		[121]
50-2700		[29]
tuaries		
6-892		[122]
2.3	0.042	[66]
3.2-32	0.032-0.16	[66]
200-3900		[123]
110-12000		[59]
14-185		[75]
< 822		[124]
594-1152		[76]
30-1764000		[125]
380-29600		[126]
54	0.08	[66]
83-2100	0.055-0.62	[66]
30-1756		[52]
1000-27000		[127]
100-9800		[128]
144-1192	0.80-4.81	[30]
100-2100	0.2-0.8	[83]
20 (1-219)	0.078 (< 0.49)	[82]
11	0.16	[66]
3.6-390		[129]
3-267	0.14-1.48	[130]
40-106	0.1-1.6	[131]
78 (2-260)	2.2 (0.78-2.34)	[88]
113-239		[132]
	20-127 38-790 10-410 50-2700 tuaries 6-892 2.3 3.2-32 200-3900 110-12000 14-185 < 822 594-1152 30-1764000 380-29600 54 83-2100 30-1756 1000-27000 100-9800 144-1192 100-2100 20 (1-219) 11 3.6-390 3-267 40-106 78 (2-260)	$\begin{array}{c c c c c c c c c c c c c c c c c c c $

taking into regard its relatively high concentration in water environment and co-migration with mercury in water [57]. Monomethylmercury is relatively stable and water-soluble, therefore it is absorbed by organisms and concentrated in the trophic chain, while dimethylmercury evaporates into the atmosphere where it can be decompose to methane and elemental mercury by photolysis or oxidised by hydroxyl radical [9, 51]:

$$(CH_3)_2Hg \twoheadrightarrow Hg^\circ + 2CH_3 \tag{3}$$

$$(CH)_2Hg + OH - \gg CH_3HgOH + CH_3$$
(4)

#### **Reduction/Oxidation Reactions**

The processes controlling conversion of elemental mercury into its ionic or organic forms determine the amount of mercury in the elemental state so the rate of mercury evaporation and the amount of total mercury remaining in water [4, 10, 60]. The reduction of mercury in ionic form Hg(II) to elemental mercury can proceed by biological and chemical processes. On the basis of experimental data some authors indicate the biological processes (with involvement of bacteria from the genus Pseudomonas and other microorganisms) as the most important, while others suggest that photochemical reactions play the dominant role [10, 21, 61-63]. According to other authors Hg(II) can be reduced by humic substances under the influence of solar radiation or without this radiation [57, 62-64]. The mechanisms of these conversions have not been well recognised yet.

As far as the process of oxidation of Hg° to Hg(II) is concerned, till recently it was considered negligible or not occurring in natural waters; however, in view of new data the process is possible, especially in marine water containing chlorine ions [61]. The oxidation processes are believed to be more important in waters close to shores where the concentration of suspension is the highest. In waters with good access to oxygen elemental mercury can be relatively quickly oxidised by different reagents - mainly oxygen but also nitrates, nitrites, iron hydroxides, iron phosphates, sulphates, sulphur, carbon dioxide [9]

#### Demethylation of Mercury

Demethylation of mercury-organic compounds is an important process taking place in natural waters. Demethylation of methyl-mercury, stable in water environments, can proceed by a biological route (through microorganisms) and a photochemical route (as a result of irradiation) [51]. Biological demethylation of mercury is a slow process and in contrast to methylation is most effective in aerobic conditions (Gilmour & Henry, 1991 vide [9]). The results reported by Matilainen & Verta [33] have indicated that demethylation takes place only with the involvement of microorganisms, because of a great influence of decreasing temperature on the rate of demethylation and cessation of demethylation in sterilised samples of water. The same authors reported that the rate of demethylation in the surface lake water was < 13.2% daily. The process of demethylation demands hydrolysis of mercury-carbon bond accompanying the formation of  $Hg^{2+}$  and methane. Then  $Hg^{2+}$  is reduced to volatile elementary mercury and released to the atmosphere where it undergoes further conversions [9]:

$$CH_3Hg^+ - WCH_3 + Hg^{2+}$$
 (5)

$$Hg^{2+} \rightarrow Hg^{\circ}$$
 (6)

#### Speciation of Mercury in Bottom Sediments

A good indicator of water contamination with mercury is its content in bottom sediments, which can store large amounts of this metal and be a site of its many conversions [9, 20, 65]. There are significant differences in the concentration of mercury in bottom sediments of different origin (Table 5). Very high concentrations have been found in sediments from the water reservoirs in polluted regions and from estuaries of some rivers.

It is assumed that a natural concentration of total mercury in bottom sediments varies from 10 to 200 ng/g dry mass [20]. Kannan & Falandysz [66] suggested that a ratio of the concentration of total mercury to that of methylmercury could be used as an index describing the pollution status of a given reservoir. In Gdansk Bay methylmercury, occurring at a mean concentration of 0.65 ng/g dry mass, made up less than 0.5% of total mercury [66]. According to the same authors, in non-polluted regions this index takes values lower than 1; for Gdansk Bay this was 0.02 to 2.27. From among the samples studied the highest index found was for bottom sediments of the harbour in Hamburg and the rivers Elba and Mulde, for which the index varied from 2.5 to 9.7 [5, 67].

The high affinity of mercury ions to sulphides determines the chemistry of mercury in anaerobic regions of waters and bottom sediments. In anaerobic conditions, the dominant mercury species are: HgS, HgS<sub>2</sub>H<sub>2</sub>, HgS<sub>2</sub>H<sup>-</sup> and  $HgS_2^{2-}$  (Fig. 4), whereas from among the mercuryorganic species the most important is CH<sub>3</sub>HgS<sup>-</sup> (Fig. 3). Mercury sulphide HgS is hardly water-soluble, is deposited in bottom sediments, and determines the solubility of Hg(II) in anaerobic waters. It seems that HgS deposition accompanies organic matter or iron oxide molecules [18]. The solubility of HgS can increase in the presence of increasing amounts of sulphide ions in the water (Fig. 4), appropriate to form soluble sulphide complexes with mercury. This property has a key importance at high concentrations of dissolved mercury noted in anaerobic regions of water ecosystems [20]. For example, in the bottom sediments from polluted Tokuyama Bay in Japan, organic species of mercury were not detected. Of the total mercury present at concentrations of 10,900 to 22,200 ng/g dry mass, the majority was bound to sulphides 50-90%, the rest occurred in the form of free ions and elemental mercury [68]. Although HgS is strongly bound to sediment, it can be partly dissolved as a result of conversions with the use of bacteria or under the effect of oxidising conditions (e.g. due to bioturbation) [18, 69-71]. The inflow of mercury from deeper layers of the sediment to near-surface layers, as a result of molecular diffusion, was estimated as 3% of the mercury inflow

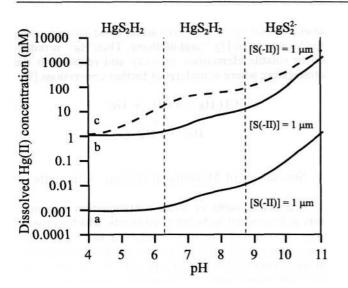
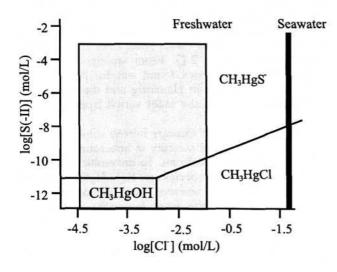


Fig. 3. Calculated dissolved Hg(II) concentrations at equilibrium with HgS in the presence of added sulphides, (a) and (b): no elemental sulphur is present, (c): the solution is at equilibrium with  $S^{\circ}$  [10].

Fig. 4. Dominance diagram of hydroxo-, chloro and sulphide complexes of methylmercury at pH = 7 as a function of chloride and total sulphide concentration [10].



from water to the sediment by Gobeil & Cossa [71], and as 7% - by Gagnon et al., [72]. According to Rutgers van der Loeff et al., [70], the size of redeposition of mercury from the sediment to water taking place thanks to the activity of benthos organisms was from 2 to 10 times greater than the redeposition from the sediment to water as a result of diffusion. It seems that the surface layer of the bottom sediments in which there are oxidising conditions make a geochemical barrier for diffusion of methylmercury from the near surface layer of the bottom sediments with reducing conditions to bulk water [18, 72].

## Conclusion

Mercury is one of the most hazardous contaminants occurring in aquatic environments, but its behaviour in hydro-systems strongly depends on the actual kind of chemical species present. Species distribution and transformation processes in natural aquatic systems are controlled by various physical, chemical and biological factors. Depending on the environmental condition, mercury species may be converted to very toxic forms (e.g. methylmercury and dimethylmercury). Recent development of analytical methods of determining mercury species in the natural environment has stimulated much effort towards the recognition of the behaviour of mercury species in the natural environment. Despite considerable literature devoted to this subject, the behaviour of mercury, processes of its conversion and distribution in natural aquatic environments are still poorly recognised.

#### References

- KURLAND L.T., FARO S.M., SEIDLER H. Minamata disease. The outbreak of neurological disorder in Min amata, Japan and its relationship to the ingestion of sea food contaminated by mercuric compounds. World Neurol ogy 1, 370, 1960.
- TAKIZAWA Y. Understanding Minamata disease and strategies to prevent further environmental contamination by methylmercury. Water Sci. Technol. 42, 139, 2000.
- KABATA-PENDIAS A., PENDIAS H. Biogeochemia pierwiastkow sladowych. PWN, Warszawa 1999.
- ULLRICH S.M., TANTON T.W., ABDRASHITOWA S.A. Mercury in the aquatic environment: a review of fac tors affecting methylation. Critical Reviews in Environ. Sci. Technol. 31, 241, 2001.
- 5. WILKEN R.D. Mercury analysis: a special example of spe cies analysis. Fresenius J. Anal. Chem. **342**, 795, **1992**.
- PUK R., WEBER J.H. Determination of mercury(II), monomethylmercury cation, dimethylmercury and diethylmercury by hydride generation, cryogenic trapping and atomic absorption spectrometric detection. Analytica Chim. Acta. 292, 175, 1994.
- TYLER G. Mercury in soil distribution, speciation and biological effect. Nordic Council of Ministers. Copenhagen, 1992.
- LINDQVIST O., JERNELOV A., JOHANSSON K, RODHE H. Mercury in Swedish Environment: Global and Local Sources. Swedish Environmental Protection Board report no. 1816. Swedish Environmental Protection Board, Stockholm, Sweden 1984.
- 9. STEIN E.D, COHEN Y., WINER A.M. Environmental distribution and transformation of mercury compounds. Critical Review in Environ. Sci. Technol. **26**, **1**, **1996**.
- MOREL F.M.M., KRAEPIEL A.M.L., AMYOT M. The chemical cycle and bioaccumulation of mercury. Annu. Rev. Ecol. Syst. 29, 543, 1998.
- BMEPC Baltic Marine Environment Protection Commis sion. Progress reports on cadmium, mercury, cooper and zinc. Baltic Sea Environment Proceedings, 24. Helsinki Commission 1987.
- PEMPKOWIAK J., COSSA D., SIKORA A., SANJUAN J. Mercury in water and sediments of the southern Baltic Sea. Sci. Total Environ. 213, 185, 1998.

- 13. BONZONGO J.C.J., LYONS W.B, WARWICK J.J., FAGNELLI J., HORVAT M, HINES M.E., LECHLER P.J., MILLER J. Aqueous geochemistry of mercury in three river systems impacted by mining activities. In: 11<sup>lh</sup> Annual International Conference on Heavy Metals in the Environ ment. Nriagu J.O. (ed.). University of Michigan, School of Public Health, Ann Arbor, MI, 2000.
- 14. GRAY J.E., THEODORAKOS P.M., BAILEY E.A, TURNER R.R. Distribution, speciation, and transport of mercury in stream-sediment, stream-water, and fish col lected near abandoned mercury mines in southwestern Alaska, USA. Sci. Total Environ. 260, 21, 2000.
- HAMASAKI T., NAGASE H., YOSHIOKA Y, SATO T. Formation, distribution, and ecotoxicology of methylmetals of tin, mercury, and arsenic in the environment. Critical Review in Environ. Sci. Technol. 25, 45, 1995.
- 16. HURLEY J.P., ROLFHUS K.R., COWELL S.E., SHAFER M.M., HUGHES P.E. Influences of watershed characteristic on total and methyl mercury levels in Lake Michigan tributaries. In: 11<sup>th</sup> Annual International Confer ence on Heavy Metals in the Environment. Nriagu J.O. (ed.). University of Michigan, School of Public Health, Ann Arbor, MI, **2000**.
- BECKER D.S., BIGHAM G.N. Distribution of mercury in the aquatic food web of Onondaga Lake, New York. Water, Air & Soil Pollut. 80, 563, 1995.
- GAGNON C, PELLETIER E., MUCCI A. Behavior of anthropogenic mercury in coastal marine sediments. Mar. Chem. 59, 159, 1997.
- TURNER D.R. Speciation cycling of arsenic, cadmium, lead and mercury in natural waters. In: Lead, Mercury, Cadmium in the Environment, Huthinson T.W., Meema K.M. (eds.), Chichester, 1987.
- FERGUSSON J.E. The heavy elements chemistry, envi ronmental impact and health effects. Pergamon Press 1990.
- 21. GALVIN R.M. Occurrence of metals in waters: an over view. Water S.A. 22, 7, 1996.
- 22. SORENSON J.A, GLASS G.E., SCHMIDT K.W., HUBER J.K., RAPP G.R. Jr. Airborne mercury deposition and watershed characteristic in relation to mercury concen tration in water, sediments, plankton, and fish of eighty northern Minnesota lakes. Environ. Sci. Technol. 24, 1716, 1990.
- COSSA D., COQUERY M, GOBEIL C, MARTIN J.-M. Mercury fluxes at the ocean margins. In: Global and Re gional Mercury Cycles: Sources, Fluxes and Mass Balances. Baeyens W. at al. (eds.), Kluwar Academic Publishers, pp 229-247, 1996.
- COQUERY M., COSSA D. Mercury speciation in surface waters of the North Sea. Neth. J. Sea Res. 34, 245, 1995.
- 25. MARINS R.V., LACERDA L.D, PARAQUETTI H.H.M. Dissolved reactive mercury speciation in water profiles from Sepetiba Bay, Brazil. In: 11<sup>th</sup> Annual International Confer ence on Heavy Metals in the Environment. Nriagu J.O. (ed.). University of Michigan, School of Public Health, Ann Arbor, MI, **2000.**
- BRYAN G.W., LANGSTON W.J. Bioavailability, accumu lation and effects of heavy metals with special reference to United Kingdom estuaries: a review. Environ. Pollut. 76, 89, 1992.
- 27. LE ROUX S.M., TURNER A, MILLWARD G.E., EB-DON L, APPRIOU P. Partitioning of mercury onto sus pended sediments in estuaries. J. Environ. Monit. **3**, 37, **2001.**

- BJORKLUND I, BORG H, JOHANSON K. Mercury in Swedish Lakes - its regional distribution and causes. Ambio 13, 118, 1984.
- DEGETTO S., SCHINTU M, CONTU A., SBRIG-NADELLO G. Santa Gilla Lagoon (Italy): a mercury sedi ment pollution case study. Contamination assessment and restoration of the site. Sci. Total Environ. 204, 49, 1997.
- MUHAYA B.B.M., LEERMAKERS M., BAEYENS W. Total mercury and methylmercury in sediments and in the polychaete *Nereis diversicolor* at Groot Buitenschoor (Sheldt Estuary, Belgium). Water, Air & Soil Pollut. 94, 109, 1997.
- BOSZKE L., FALANDYSZ J. Rtec w warstwie powierzchniowej oraz rdzeniach osadow dennych z Zatoki Puckiej. Brom. Chem. Toksykol. 32, 69, 1999.
- 32. BOSZKE L. Rtec w ekosystemie Zatoki Puckiej: stezenia w skladnikach biotycznych i abiotycznych, rozmieszczenie przestrzenne, ocena zrodel, historia zanieczyszczenia i bilans. Praca doktorska. Uniwersytet Gdanski. Gdansk, 1999.
- MATILAINEN T., VERTA M. Mercury methylation and demethylation in aerobic surface waters. Can. J. Fish. Aquat. Sci. 52, 1597, 1995.
- GUENTZEL J.L., POWELL R.T, LANDING W.M., MA SON R.P. Mercury associated with colloidal material in estuarine and open-ocean environment. Mar. Chem. 55, 177, 1996.
- MEILI M. Mercury in lakes and rivers. Metal Ions Biol. Syst. 34, 21, 1997.
- WALLSCHLAGER D, DESAI M.V.M., WILKEN R.D. The role humic substances in the aqueous mobilization of mercury from contaminated floodplain soils. Water, Air & Soil Pollut. 90, 507, 1996.
- WALLSCHLAGER D, DESAI M.V.M., SPRENGLER M., WINDMOLLER C.C, WILKEN R.D. How humic substances dominate mercury geochemistry in con taminated floodplain soils and sediments. J. Environ. Qual. 27, 1044, 1998.
- LOUX N.T. An assessment of mercury-species-dependent binding with natural organic carbon. Chem. Speciation Bi oavailability. 10, 127, 1998.
- SCHWESIG D., MATZNER E. Pools and fluxes of mer cury and methylmercury in two forested catchments in Ger many. Sci. Total Environ. 260, 213, 2000.
- SCHWESIG D., MATZNER E. Dynamics of mercury and methylmercury in forest floor and runof of forested water shed in Central Europe. Biogeochem. 53, 181, 2001.
- BIGHAM G.N., VANDAL G.M. A drainage basin per spective of mercury transport and bioaccumulation: Onon daga Lake, New York. NeuroToxicology 17, 279, 1996.
- 42. NEUMANN K., LYONS W.B., ROSI E.J., MEYER J.L. Hg and other trace metals in water, suspended matter, and biota in the Chattahoochee River. In: 11<sup>th</sup> Annual Interna tional Conference on Heavy Metals in the Environment. Nriagu J.O. (ed.). University of Michigan, School of Public Health, Ann Arbor, MI, **2000**.
- ST LOUIS V.L, RUDD J.W.M., KELLY C.A., BEATY K.G., FLETT R.J., ROULET N.T. Production and loss methylmercury from boreal forest catchments containing different types of wetland. Environ. Sci. Tech. 30, 2719, 1996
- MASON R.P, LAWSON N.M., SULLIVAN K.A. Atmos pheric deposition to the Chesapeake Bay watershed - re gional and local sources. Atmos. Environ. 21, 3531, 1997.

- 45. ROSSMANN R. Mercury in and fluxes to lake Michigan surficial sediments. In: 11<sup>th</sup> Annual International Confer ence on Heavy Metals in the Environment. Nriagu J.O. (ed.). University of Michigan, School of Public Health, Ann Arbor, MI, **2000.**
- RUDD J.W.M. Sources of methyl mercury to freshwater ecosystems: a review. Water, Air & Soil Pollut. 80, 697, 1995
- 47. PORVARI P., VERTA M. The role of catchment as a methylmercury source to lakes. In: 11<sup>th</sup> Annual Interna tional Conference on Heavy Metals in the Environment. Nriagu J.O. (ed.). University of Michigan, School of Public Health, Ann Arbor, MI, **2000.**
- WESTCOTT K., KALFF J. Environmental factors affect ing methyl mercury accumulation in zooplankton. Can. J. Fisch. Aqua. Sci. 53, 2221, 1996.
- PETTERSSON C, BISHOP K, LEE Y.-H, ALLARD B. Relations between organic carbon and methylmercury in humic rich surface waters from Svartberget catchment in northern Sweden. Water, Air & Soil Pollut. 80, 971, 1995.
- REGNELL O, TUNLID A., EWALD G, SANGFORS O. Methyl mercury production in freshwater microcosms af fected by dissolved oxygen levels: role of cobolamin and microbial community composition. Can. J. Fish. Aquat. Sci. 53, 1535, 1996.
- HOBMAN J.L., WILSON J.R., BROWN N.L. Microbial Mercury Reduction. In: Environmental Microbe-Metal In teraction. Lovley D. R. (ed.) ASM Press, Washington 2000.
- LEERMAKERS M, ELSKENS M., PANUTRAKUL S., MONTENY F., BAEYENS W. Geochemistry of mercury in an intertidal flat of the Scheldt Estuary. Netherlands J. Aqua. Ecol. 27, 267, 1993.
- MATILAINEN T. Involvent of bacteria in methylmercury formation in anaerobic lake waters. Water, Air & Soil Pol lut. 80, 757, 1995.
- BENOIT J.M., GILMOUR C.C., MASON R.P. Aspects of bioavailability of mercury for methylation in pure cultures of Desulfobulbus propionicus. Appl. Environ. Microbiol. 67, 51, 2001.
- COMPEAU G., BARTHA R. Effects of sea salt on the formation and stability of methylmercury. Bull. Environ. Toxicol. 31, 486, 1983.
- CRAIG P., MORETON P.A. Total mercury, methyl mer cury and sulphide levels in British estuarine sediments-III. Water Res. 20, 1111, 1986.
- 57. WEBER J.H. Review of possible paths for abiotic methyla tion of mercury(II) in aquatic environment. Chemosphere **51**, 2063, **1993**.
- CERATTI G, BERNHARD M., WEBER J.H. Model re actions for abiotic mercury (II) methylation: kinetics of methylation of mercury (II) by mono-, di- and tri- methyltin in seawater. Applied Organomet. Chem. 6, 587, 1992.
- EBINGHAUS R., HINTELMAN H., WILKEN R.D. Mer cury-cycling in surface waters and in the atmosphere - spe cies analysis for the investigation of transformation and transport properties of mercury. Fresenius J. Anal. Chem. 350, 21, 1994.
- LOUX N.T. Dial temperature effects on the exchange of elemental mercury between the atmosphere and underlying waters. Environ. Toxicol. Chem. 19, 1191, 2000.
- AMYOT M., GILL G.A., MOREL F.M.M. Production and loss of dissolved gaseous mercury in coastal seawater. En viron. Sci. Technol. 31, 3606, 1997.

- COSTA M., LISS P. Photoreduction of mercury in seawater and its possible implications for Hg° air-sea fluxes. Mar. Chem. 68, 87, 1999.
- COSTA M., LISS P. Photoreduction and evolution of mer cury from seawater. Sci. Total Environ. 161, 125, 2000.
- LIU J.H, WANG W.H, PENG A. The photochemical re duction of divalent mercury and methyl-mercury. J. En viron. Sci. Health Part A-Toxic/Hazardous Subst. Eng. 35, 1859, 2000.
- BOUDOU A., RIBEYRE F. Contamination of aquatic biocenoses by mercury compounds: an experimental ecotoxicological approach. In: Aquatic toxicology. Nriagu J.O. (ed.), Wiley J. & Sons, New York, pp 73-116, **1983**.
- KANNAN K, FALANDYSZ J. Speciation and concentra tions of mercury in certain coastal marine sediments. Water, Air & Soil Pollut. 103, 129, 1998.
- 67. WILKEN R.D., HINTELMANN H. Mercury and methyl mercury in sediments and suspended particles from the river Elbe, North Germany. Water, Air & Soil Pollut. 56, 427, 1991.
- NAKANISHI H., UKITA M., SERINE M., MURAKAMI S. Mercury pollution in Tokuyama Bay. Hydrobiology 176/177,197,1989.
- LUOMA S.N., DAVIS J.A. Requirements for modelling trace metal partitioning in oxidized estuarine sediments. Mar. Chem. 12, 159, 1983.
- RUTGERS VAN DER LOEFF M.M., ANDERSON L.G., HALL P.O.J., IVERFELDT A., JOSEFSON A.B., SUNBY B., WESTERLUND S.F.G. The asphyxiation technique: An approach to distinguishing between molecu lar diffusion and biologically mediated transport at the sedi ment-water interface. Limnol. Oceanogr. 29, 675, 1984.
- GOBEIL C, COSSA D. Mercury in sediments and sedi ment pore water in Laurentian Trough. Can. J. Fish. Aquat. Sci. 50, 1794, 1993.
- GAGNON C, PELLETIER E., MUCCI A, FIT ZGERALD W.F. Daigenetic behavior of methylmercury in organic-rich coastal sediments. Limnol. Oceanogr. 41, 428, 1996.
- 73. SCHMIDT D. Mercury in Baltic and North Sea waters. Water, Air & Soil Pollut. **62**, 43, **1991**.
- NASFI F.H. Total mercury content of sea water on the Tunisian Shore. Fresenius Environ. Bull. 4, 161, 1995.
- 75. SCHUHMACHER M., DOMINGO J.L., LOBET J.M., CORBELLA J. Evaluation of effect of temperature, pH, and bioproduction on Hg concentration in sediments, water, molluscs and algae of the Delta of the Ebro River. Sci. Total Environ. Supplement. 117, **1993.**
- AY AS Z., KOLANKAYA D. Accumulation of some heavy metals in various environments and organisms at Goksu Delta, 1991-1993, Turkiye. Bull. Environ. Contam. Toxicol. 56, 65, 1996.
- VANDAL G.M., FITZGERALD W.F. A preliminary mer cury budget for Narragansett Bay (Rhode Island, USA). Water, Air & Soil Pollut. 80, 679, 1995.
- 78. HURLEY J.P., BENOIT J.M., BABIARZ C.L., SHAF FER M.M., ANDREAN A.W, SULLIVAN J.R., HAM MOND R., WEBB D.A. Influences of watershed character istics on mercury levels in Wisconsin rivers. Environ. Sci. Tech. 29, 1867, 1995.
- 79. ROULET M, LUCOTTE M, CANUEL R, RHEAULT J., TRAN S., DE FREITOS GOG Y.G., FERALLA N., SOUSA DA VALE R., SOUSA PASSOS C.J., DE JESUS

DA SILVA EL, MARGLER D., AMORIM M. Distribution and partition of total mercury in waters of the Tapajos River Basin, Brasilian Amazon. Sci. Total Environ. **213**, 203, **1998**.

- QUEMARAIS R., COSSA D., RONDEAU B, PHAM T.T., FARTIN B. Mercury distribution in relation to iron and manganese in the waters of St Lawrence River. Sci. Total Environ. 213, 193, 1998.
- COSSA D., GOBEIL C. Mercury speciation in the St Law rence Estuary. Can. J. Fish. Aquat. Sci. 57, 138, 2000.
- KANNAN K., SMITH R.G., LEE R.F., WINDON H.L., HEIMULLER P.T., MACAULCY J.M., SUMMERS J.K. Distribution of total mercury and methyl mercury in water, sediment, and fish from South Florida estuaries. Arch. En viron. Contam. Toxicol. 34, 109, 1998.
- 83. SOUTHWORTH G.R., BOGLE M.A., TURNER R.R. Mercury and methylmercury relationships in contaminated streams in the southeastern USA. In: 11<sup>th</sup> Annual Interna tional Conference on Heavy Metals in the Environment. Nriagu J.O. (ed.). University of Michigan, School of Public Health, Ann Arbor, MI, 2000.
- 84. TSENG CM., AMOUROUX D., DONARD O.F.X. Biogeochemistry of mercury in three European estuaries (Gironde, Scheldt and Rhine). In: 11<sup>th</sup> Annual Interna tional Conference on Heavy Metals in the Environment. Nriagu J.O. (ed.). University of Michigan, School of Public Health, Ann Arbor, MI, 2000.
- HURLEY J.P., COWEL S.E., SHAFER M.M., HUGHES P.H. Tributary loading of mercury to Lake Michigan. Im portance of seasonal events and phase partitioning. Sci. To tal Environ. 213, 129, 1998.
- SULLIVAN K., MASON R.P. The concentration and dis tribution of mercury in Lake Michigan. Sci. Total Environ. 213, 213, 1998.
- MASON P.R., SULLIVAN K.A. Mercury in the water col umn of Lake Michigan In: 11<sup>th</sup> Annual International Con ference on Heavy Metals in the Environment. Nriagu J.O. (ed.). University of Michigan, School of Public Health, Ann Arbor, MI, **2000**.
- ZHANG X., RYGWELSKI K.R. A modeling framework for mercury cycling in Lake Michigan. In: 11<sup>th</sup> Annual Inter national Conference on Heavy Metals in the Environment. Nriagu J.O. (ed.). University of Michigan, School of Public Health, Ann Arbor, MI, 2000.
- PLOURDE Y., LUCOTTE M, PICHET P. Contribution of suspended particulate matter and zooplankton to MeHg contamination of the food chain in midnorthern Quebec (Canada) reservoirs. Can. J. Fish. Aquat. Sci. 54, 821, 1997.
- MARINS R.V., LACERDA L.D. Tidal remobilisation of mercury from mud flat acidic pore waters. In: II"<sup>1</sup> Annual International Conference on Heavy Metals in the Environ ment. Nriagu J.O. (ed.). University of Michigan, School of Public Health, Ann Arbor, MI, 2000.
- HULTBERG H., IVERFELDT A., LEE Y.H. Methylmer cury input/output and accumulation from forested catch ments and critical loads for lakes in southwestern Sweden. In: Mercury Pollution: Integration and Synthesis. Watras C.J., Huckabee J.W. (eds.), Lewis Publishers, Chelsea, pp 313-322, 1994.
- 92. EBINGHAUS R., KOCK H.H., JENNINGS S.G., McCARTIN P., ORREN M.J. Measurement of atmos pheric mercury concentration in northwestern and central Europe - comparison of experimental data and model re sults. Atmos. Environ. 29, 3333, 1995.

- IVERFELDT A. Occurrence and turnover of atmospheric mercury over the Nordic countries. Water, Air & Soil Pollut. 56, 251, 1991.
- ST LOUIS V.L., RUDD J.W.M., KELLY C.A., BARRIE L.A. Wet deposition of methyl mercury in northwestern Ontario compared to other locations. Water, Air & Soil Pollut. 80, 405, 1995.
- POISSANT L., PILOTE M. Mercury concentrations in single event precipitation in southern Quebec. Sci Total En viron. 213, 65, 1998.
- DVONCH J.T., GRANEY J.R., MARSIK F.J., KEELER G.J., STEVENS R.K. An investigation of source-receptor relationship for mercury in south Florida using event pre cipitation data. Sci. Total Environ. 213, 95, 1998.
- HOYER M., BURKE J., KEELER G. Atmospheric sour ces, transport and deposition of mercury in Michigan: two years of event precipitation. Water, Air & Soil Pollut. 80, 199, 1995.
- GUENTZEL J.L., LANDING W.M., GILL G.A., POLL-MAN CD. Atmospheric deposition of mercury in Florida: the fams project (1992-1994). Water, Air & Soil Pollut. 80, 393, 1995.
- 99. HOFF R.M., STRACHAN W.M.J., SWEET C.W., CHAN C.H., SHACKLETON M., BIDLEMAN T.F., BRICE K.A., BURNISTON D.A., CUSSION S., GATZ D.F., HARLIN K., SCHROEDER W.H. Atmospheric deposi tion of toxic chemicals to the Great Lakes: a review of data through 1994. Atmos. Environ. **30**, 3505, **1996**.
- LAMBORG C.H., FITZGERALD W.F., VANDAL G.M., ROLFHUS K.R. Atmospheric mercury in northern Wis consin: sources and species. Water, Air & Pollut. 80, 189, 1995.
- MORRISON K.A., KUHN E.S., WATRAS C.J. Compari son of three methods of estimation atmospheric mercury deposition. Environ. Sci. Technol. 29, 571, 1995.
- 102. FITZGERALD W.F., MASON R.P., VANDAL G.M. At mospheric cycling and air-water exchange of mercury over mid-continental lacustrine regions. Water, Air & Soil Pol lut. 56, 745, 1991.
- 103. HAINES T.A., ABBOTT C.C., RYAN J.M. Mercury pools and pathways in two small stream watersheds in Maine, and the role of watershed acidification. In: The International Conference on Mercury as a Global Pollutant. Whistler, British Columbia, **1994**.
- NAKAGAWA R. Studies on the levels in atmospheric con centrations of mercury in Japan. Chemosphere 31, 2669, 1995.
- BORG H., JONSSON P. Large-scale metal distributionn in Baltic Sea sediments. Mar. Pollut. Bull. 32, 8, 1996.
- LEIVUORI M., NIEMISTO L. Sedimentation of trace metals in the Gulf of Bothnian. Chemosphere 31, 3839, 1995.
- HERMANSON M.H. Antropogenic mercury deposition to arctic lake sediments. Water, Air & Soil Pollut. 101, 309, 1998.
- 108. HENRY E.A., DODGE-MURPHY L.J., BIGHAM G.N., KLEIN S.M., GILMOUR C.C. Total mercury and methylmercury mass balance in an alkaline hypereutrophic urban lake (Onondaga Lake, New York). Water, Air & Soil Pol lut. 80, 509, 1995.
- 109. HURLEY J.P., WATRAS C.J., BLOOM N.S. Distribution and flux particulate mercury in four stratified seepage lakes. In: Mercury Pollution: Integration and Synthesis. Watras CJ, Huckabee JW (eds.), Lewis Publishers, Chelsea, pp 69-82, **1994**.

- ENGSTROM D.R., SWAIN E.B. Recent declines in at mospheric mercury deposition in the upper Midwest. En viron. Sci. Technol. **31**, 960, 1997.
- 111. VERTA M., MATILAINEN T., PORVARI P., NIEMI M, UUSIRAUVA M, BLOOM N.S. Methylmercury sources in boreal lake ecosystems. In: Mercury Pollution: Integra tion and Synthesis. Watras C.J., Huckabee J.W. (eds.), Lewis Publishers, Chelsea, pp. 119-136, **1994.**
- 112. JOHANSSON K, AASTRUP M., ANDERSSON A., BRINGMARK L., IVERFELDT A. Mercury in Swedish forests and soils and waters-assessment of critical load. Water, Air & Soil Pollut. 56, 267, 1991.
- 113. AASTRUP M., JOHNSON J, BRINGMARK E., BRING MARK L., IVERFELDT A. Occurrence and transport of mercury within a small catchment area. Water, Air & Soil Pollut. 56, 155, 1991.
- 114. BRZEZINSKA A., TRZOSINSKA A, ZMIJEWSKA W., WODKIEWICZ L. Trace metals in suspended matter and surficial bottom sediments from the Southern Baltic. Oceanologia 18, 59, 1984.
- 115. FALANDYSZ J, DUBRAWSKI R., BOLALEK J. Zawartosc rt?ci w osadach dennych z Zatoki Puckiej i Gdanskiej. Bromat. Chem. Toksykol. **26**, 29, **1993.**
- 116. OJAVER E. (ed.). Mercury content in the ecosystem of the Gulf of Riga. In: Ecosystem of the Gulf of Riga between 1920 and 1990. pp 80-103, Tallin **1995.**
- 117. HERUT B., HORNUNG H, KRESS N. Mercury, lead, copper, zinc and iron in shallow sediments of Haifa Bay, Israel. Fresenius Environ. Bull. **3**, 147, **1994.**
- 118. HAYNES D., TOOHEY D., CLARKE D, MARNEY D. Temporal and spatial variation in concentrations of trace metals in coastal sediments from the Ninety Mile Beach, Victoria, Australia. Mar. Pollut. Bull. 30, 414, 1995.
- 119. BARGHIGIANI C, RISTORI T. Preliminary results on the role of rivers in total Hg concentrations in marine sedi ments and benthic organisms of coastal area of Italy. Water, Air & Soil Pollut. 80, 1017, 1995.
- 120. ISKANDER F.Y., VEGA-CARRILLO H.R., ACUNA E.M. Determination of mercury and other elements in La Zacatecana Dam sediment in Mexico. Sci. Total Environ. 148, 45, 1994.
- **121.** AMIN O., FERRER L., MARCOVECCHIO J. Heavy metal concentrations in littoral sediments from the Baeagle Channel, Tierra Del Fuego, Argentina. Environ. Monit. As sess. **41**, 219, **1996**.

- 122. FALANDYSZ J., STEPNOWSKI P. Mercury in surface sediments and sediment cores of the Motlawa River. Chem. Anal. (Warsaw) 41, 1051, 1996.
- 123. HELIOS-RYBICKA E., STRZEBONSKA M. Distribution and chemical forms of heavy metals in the flood 1997 sedi ments of the upper and middle Odra River and its tribu taries, Poland. Acta Hydrochim. Hydrobiol. 27, 331, 1999.
- 124. SIEGEL F.R., SLABODA M.L., STANLEY D.J. Metal pollution loading, Manzalah Lagoon, Nile Delta, Egypt: implications for agriculture. Environ. Geology 23, 89, 1994.
- 125. JOHNSTON P., STRINGER R., FRENCH M.C., VAL-LETE J. Contamination of soils and sediments in the vicin ity of a mercury recovery plant. Bull. Environ. Contam. Toxicol. 46, 74, 1991.
- 126. BONNEVIE N.L., WENNING R.J., HUNTLEY S.L., BE-DBURY H. Distribution of inorganic compounds from three waterways in Northern New Jersey. Bull. Environ. Contam. Toxicol. 51, 672, **1993.**
- 127. WINGER P.V., LASIER P.J., GEINER H. Toxicity of sediments and pore water from Brunswick Estuary, Geor gia. Arch. Environ. Contam. Toxicol. 25, 371, 1993.
- GILLIS C.A, BONNEVIE N.L., WENNING R.J. Mercury contamination in the Newark Bay Estuary. Ecotoxicol. En viron. Saf. 35, 214, 1993.
- 129. FALANDYSZ J., KAWANO M., DANISIEWICZ D., STEPNOWSKI P, BOSZKE L., CHWIR A., GOLEBIOWSKI M., KRYSZEWSKI K. Mercury concen tration in freshwater and marine sediments from various sites in Poland. 5<sup>th</sup> Polish Conference on Analytical Chemis try, Gdansk, 3-9 September, 1995.
- 130. TREMBLAY A., LUCOTTE M, MEILI M., CLOUTIER L., PICHET P. Total and methylmercury contents of insect from boreal lakes: ecological, spatial and temporal patterns. Water Qual. Res. J. Canada 31, 851, 1996.
- 131. TREMBLAY A., LUCOTTE M. Accumulation of total mercury and methyl mercury in insect larvae of hydroelec tric reservoirs. Can. J. Fish. Aquat. Sci. 54, 832, 1997.
- 132. ALMEIDA M.G., SOUZA C.M.M., GUEDES L. Geochemical study of heavy metals in bottom sediments and border soil from lacustrine system of Cima Lake, RJ, Brazil. W: 11<sup>th</sup> Annual International Conference on Heavy Metals in the Environment. Nriagu J.O. (ed.). Uni versity of Michigan, School of Public Health, Ann Arbor, MI, **2000.**